

Baeckeins A and B, Two Novel 6-Methylflavonoids from the Roots of *Baeckea frutescens*

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A pair of novel isomeric 6-methylflavonoids, named baeckeins A (**1**) and B (**2**), were isolated from the roots of *Baeckea frutescens*. The two compounds possess a unique C₂₃ skeleton, resulting from the 6-methylation and 8-arylation of a flavonol (= 3-hydroxy-2-phenyl-4*H*-1-benzopyran-4-one) framework and the formation of an unusual lactone ring *E*. Their structures were elucidated by detailed spectroscopic analyses, including HR-ESI-MS and 1D- and 2D-NMR data (HSQC, HMBC, and ROESY). A plausible biogenetic pathway for compounds **1** and **2** is also proposed (*Scheme*).

Introduction. – *Baeckea frutescens* L. (Myrtaceae) is an aromatic low-growing shrub that ranges from Southeast Asia to Australia. It is widely used as a folk medicine in Thailand, Peninsular Malaysia, Sumatra, Borneo, Sulawesi, and New Guinea. In traditional Chinese medicine, the roots of *B. frutescens* have been used for treating rheumatism and snake bites [1]. Previous investigations of this species revealed the presence of essential oil [2], sesquiterpenes [3], phloroglucinols [4], chromones [5], flavonoids including flavanone [6], flavones, and further derivatives [7][8]. Some flavonoids exhibited strong antioxidant [9] and cytotoxic [6] properties. In our ongoing research on the flavonoids of *B. frutescens*, a pair of novel 6-methylflavonoids, baeckeins A¹⁾ (**1**) and B (**2**), were isolated from the roots of this plant (*Fig. 1*). In this article, we describe the isolation, structure elucidation, and possible biosynthetic pathway of compounds **1** and **2**.

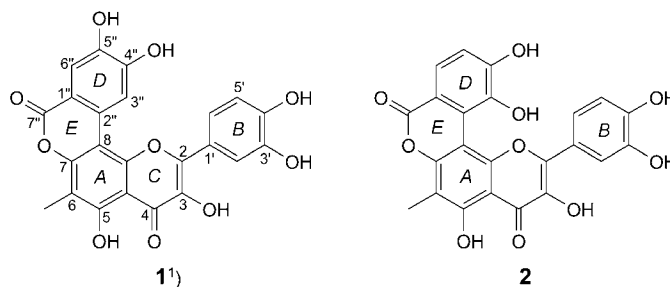


Fig. 1. *Baeckeins A (1) and B (2), isolated from Baeckea frutescens*

¹⁾ Trivial atom numbering; for systematic names, see *Esper. Part*.

Results and Discussion. – Baecklein A¹) (**1**) was obtained as a yellow amorphous powder and gave a quasimolecular-ion peak at m/z 449.0527 ($[M - H]^-$) in the HR-ESI-MS, consistent with the elemental composition C₂₃H₁₄O₁₀. Its IR spectrum showed absorption bands for OH groups (3427 cm⁻¹), two C=O groups (1710 and 1643 cm⁻¹), and aromatic functionalities (1609 and 1523 cm⁻¹). The UV spectrum exhibited maxima at 254 and 345 nm, and the positive result for the Mg/HCl reaction suggested that compound **1** was a flavonoid. The ¹H- and ¹³C-NMR spectra (Table) of **1** showed signals assignable to a flavonol (= 3-hydroxy-2-phenyl-4*H*-1-benzopyran-4-one) moiety by a typical *ABX* coupling system for H–C(2') (δ (H) 6.95 (*d*, $J = 8.5$)), H–C(5') (δ (H) 7.69 (*d*, $J = 2.2$)), and H–C(6') (δ (H) 7.48 (*dd*, $J = 8.5, 2.2$)), the signal of one strongly chelated OH group, *i.e.*, of OH–C(5) (δ (H) 13.45 (*s*)), and three characteristic C-signals, *i.e.*, of C(4) (δ (C) 176.3), C(3) (δ (C) 136.5), and C(2) (δ (C) 147.9). An additional aromatic Me signal at δ (H) 2.25 (*s*) was observed, and the Me group was located at C(6) by the HMBC cross-peaks *Me*–C(6)/C(5), C(6), and C(7) (Fig. 2). By comparison of the NMR data with those of the known compound 6-methylquercetin [9], the above mentioned signals suggested the presence of a 6-methylquercetin unit, but the absence of a H–C(8) signal revealed that the C(8) position was substituted (6-methylquercetin = 2-(3,4-dihydroxyphenyl)-3,5,7-trihydroxy-6-methyl-4*H*-1-benzopyran-4-one). Except for the signals of 6-methylquercetin, the ¹³C-NMR spectrum

Table. ¹H- and ¹³C-NMR Data (500 and 125 MHz, resp.; (D₆)DMSO) of **1** and **2**¹). δ in ppm, J in Hz.

	1		2	
	δ (H)	δ (C)	δ (H)	δ (C)
C(2)		147.9		147.9
C(3)		136.5		136.4
C(4)		176.3		176.0
C(5)		156.7		157.6
C(6)		106.7		105.6
C(7)		153.2		153.4
C(8)		99.4		98.3
C(9)		150.2		149.4
C(10)		106.3		106.1
C(1')		121.4		121.8
H–C(2')	6.95 (<i>d</i> , $J = 8.5$)	115.6	6.83 (<i>d</i> , $J = 8.5$)	115.3
C(3')		145.2		144.7
C(4')		149.4		148.3
H–C(5')	7.69 (<i>d</i> , $J = 2.2$)	116.2	7.84 (<i>d</i> , $J = 2.2$)	116.1
H–C(6')	7.48 (<i>dd</i> , $J = 8.5, 2.2$)	120.2	7.64 (<i>dd</i> , $J = 8.5, 2.2$)	120.7
C(1'')		110.8		112.5
C(2'')		126.5		119.3
H–C(3'') or C(3'')	8.30 (<i>s</i>)	112.2		140.9
C(4'')		146.1		152.9
C(5'') or H–C(5'')		153.4	7.12 (<i>d</i> , $J = 8.4$)	115.4
H–C(6'')	7.62 (<i>s</i>)	114.4	7.70 (<i>d</i> , $J = 8.4$)	122.4
C(7'')		159.1		160.0
<i>Me</i> –C(6)	2.25 (<i>s</i>)	7.5	2.25 (<i>s</i>)	7.4
OH–C(5)	13.45 (<i>s</i>)		13.50 (<i>s</i>)	

displayed seven additional signals, among which two arose from CH moieties and five from quaternary C-atoms (two of them O-bearing), and the ^1H -NMR spectra exhibited two for two aromatic H-atoms. Comprehensive analysis of the ^1H -NMR and HSQC spectra established the presence of a 4'',5''-dioxo-substituted ring *D*, characterized by $\delta(\text{C})$ 110.8 (C(1'')), 126.5 (C(2'')), 112.2 (C(3'')), 146.1 (C(4'')), 153.4 (C(5'')), and 114.4 (C(6'')) and $\delta(\text{H})$ 8.30 (*s*, H–C(3'')) and 7.62 (*s*, H–C(6'')). The unusual ester C=O signal at $\delta(\text{C})$ 159.1 (C(7'')) was consistent with the IR absorption band at 1710 cm^{-1} and suggested the presence of an aromatic lactone ring *E*, in accord with the remaining one degree of unsaturation. The HMBCs H–C(3'')/C(1''), C(4''), and C(5''), and H–C(6'')/C(2''), C(5''), and C(7'') (Fig. 2) confirmed the presence of the C_7 unit, which was connected to the 6-methylquercetin moiety *via* the C(8)–C(2'') bond as suggested by the cross-peaks H–C(3'')/C(8). The presence of the lactone ring *E* (C(7), C(8), C(2''), C(1''), and C(7'')) was confirmed by comparison of the ^{13}C -NMR data with those reported for 3,4-benzo-fused furocoumarins [10]. The NOE correlations H–C(3'')/H–C(6') and H–C(5') (Fig. 2) definitively established the structure of compound **1**.

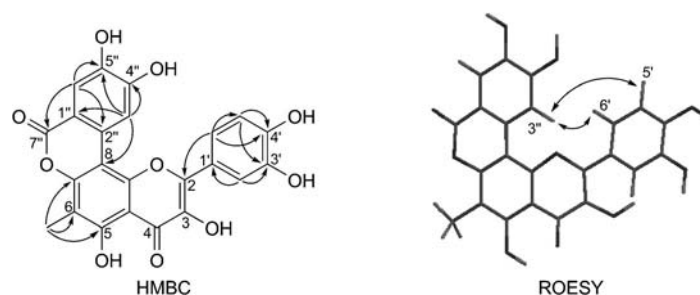


Fig. 2. Selected 2D-NMR correlations of compound **1**

Baeckein B¹) (**2**) was also obtained as a yellow amorphous powder, and possessed the molecular formula $\text{C}_{23}\text{H}_{14}\text{O}_{10}$, as established by the HR-ESI-MS (m/z 449.0498 ($[M - \text{H}]^-$), indicating an isomer of **1**. Its IR and UV spectra showed similar patterns to those of **1**. Careful inspection of the ^1H - and ^{13}C -NMR data of compound **2** (Table) revealed some structural characteristics of compound **1** and also suggested the presence of a 6-methylquercetin unit, a C_7 unit, and the lactone ring *E*. The differences between compounds **1** and **2** concerned only ring *D*. The easily visible changes were a pair of *ortho*-coupled aromatic H-atoms, *i.e.*, the signals of H–C(5'') ($\delta(\text{H})$ 7.12 (*d*, $J = 8.4$)) and H–C(6'') ($\delta(\text{H})$ 7.70 (*d*, $J = 8.4$)), instead of the two *s* at $\delta(\text{H})$ 8.30 and 7.62 (each 1 H), for **1** suggesting a 3'',4''-dioxo-substituted ring *D* in **2**, characterized by $\delta(\text{C})$ 112.5 (C(1'')), 119.3 (C(2'')), 140.9 (C(3'')), 152.9 (C(4'')), 115.4 (C(5'')), and 122.4 (C(6'')). The above conclusion was confirmed by the HMBC H–C(5'')/C(1''), C(3''), and C(4''), and H–C(6'')/C(2''), C(4''), and C(7'') and the ROESY data of compound **2** (no NOE between ring *D* and ring *B*) (Fig. 3). On the basis of these evidences, the structure of **2** was determined.

From a chemical point of view, compounds **1** and **2** possess a unique C_{23} skeleton, resulting from the 6-methylation and 8-arylation of a flavonol framework and the

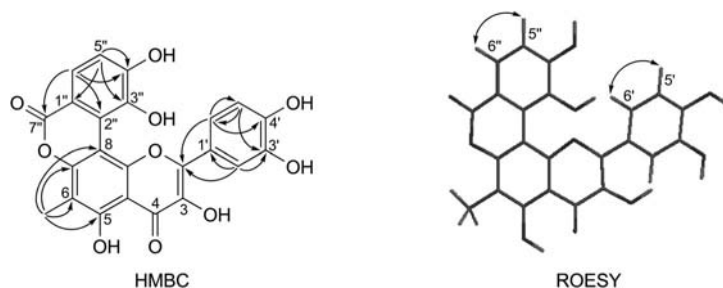


Fig. 3. Selected 2D-NMR correlations of compound **2**

formation of an unusual lactone ring *E*. C-Methylation of ring *A* is common in flavonoids from *B. frutescens* and more generally in the family Myrtaceae [9], but 8-arylflavones (except biflavonoids with a C–C connection to C(8)) are rare in nature [11][12]. A plausible biogenetic pathway for baeckeins A (**1**) and B (**2**) is proposed in the *Scheme*. The 6-Methylquercetin (**3**), which has been already isolated from *B. frutescens* [9], and protocatechuic acid (**4**), a natural phenol-derived acid which provides the *C*₇ unit, are suggested as precursors. The mechanism involves a phenolic coupling *via* a radical mechanism. Such a coupling takes place typically between *ortho*–*para*, *ortho*–*ortho*, or *para*–*para* positions [13][14]. Compounds **3** and **4** can generate free radicals **3b** and **4b** and **4c**, respectively. For compound **1**, the coupling is between *ortho*- and *para*-C-atoms (**3b** and **4c**), while two *ortho*-C-atoms (**3b** and **4b**) are coupled in the case of **2**. The lactone ring *E* can be easily formed by subsequent intramolecular esterification. Compounds **1** and **2** are, to the best of our knowledge, the first conjugates of protocatechuic acid and a flavonol leading to a unique C₂₃ skeleton.

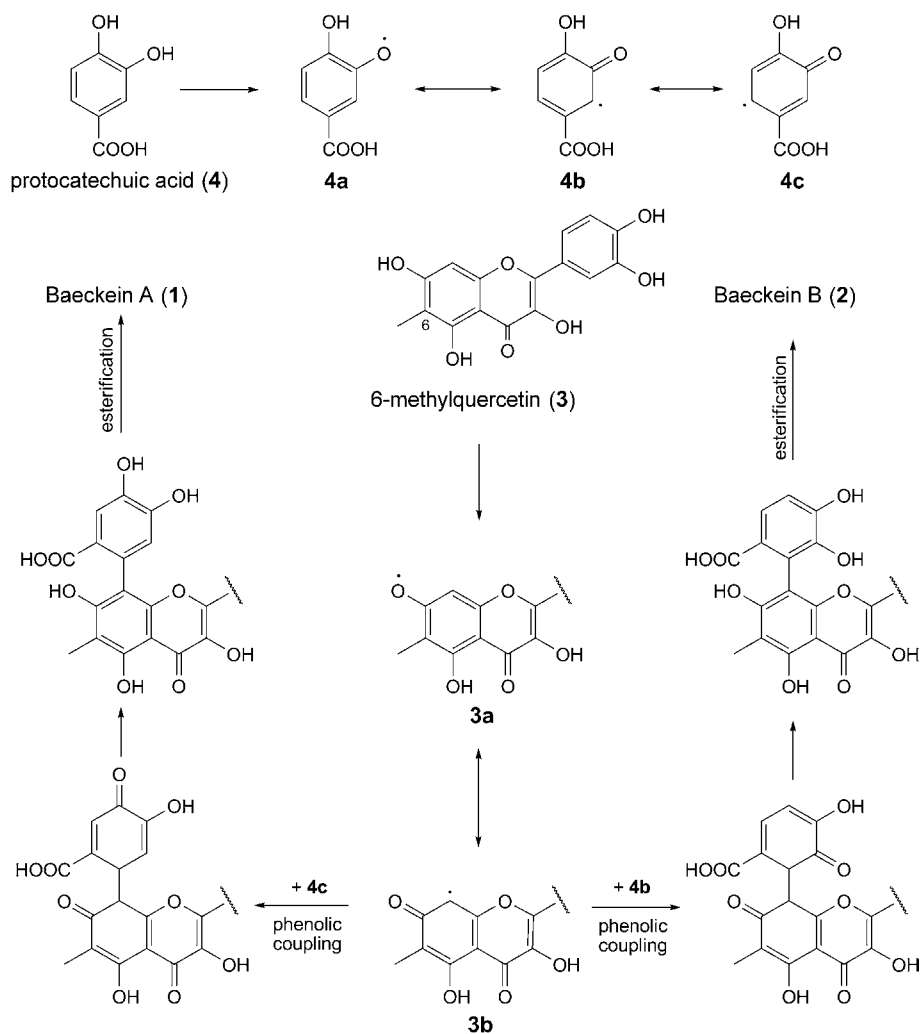
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Experimental Part

General. Column chromatography (CC): silica gel (SiO₂, 200–300 mesh; *Qingdao Haiyang Chemical Co., Ltd.*) and *Sephadex LH-20* (20–100 μm; *Pharmacia*). UV Spectra: *Shimadzu-UV-2450* UV/VIS spectrophotometer; λ_{max} (log ε) in nm. IR Spectra: *Nicolet-Impact-410* spectrometer; KBr discs; ν̄ in cm^{−1}. 1D- and 2D-NMR Spectra: *Bruker-AV-500* spectrometer; δ in ppm rel. to Me₄Si as internal standard, *J* in Hz. ESI and HR-ESI-MS: *Agilent LC/MSD TOF*; in *m/z*.

Plant Material. The roots of *B. frutescens* were collected from Nanning, Guangxi Province, P. R. China, in Nov. 2009, and identified by *Q. W.*, China Pharmaceutical University. A voucher sample (No. GS001) is kept with the Department of Chinese Material Medica Analysis, China Pharmaceutical University, Nanjing, P. R. China.

Extraction and Isolation. The dried roots of *B. frutescens* (10 kg) were extracted with 90% EtOH (3 × 30 l) at 80° for 3 h and concentrated to give 600 g of extract, which was suspended in H₂O (5 l) and partitioned successively with petroleum ether (3 × 5 l), AcOEt (10 × 5 l), and BuOH (10 × 5 l). The AcOEt-soluble part (203.6 g) was subjected to CC (CHCl₃/MeOH 1:0 → 1:1); *Fractions 1–8*. *Fr. 5* (15.8 g) was submitted to repeated CC (CHCl₃/MeOH 25:1 → 5:1), and the obtained **1** was purified by

Scheme. Plausible Biogenetic Pathway for Compounds **1** and **2**

CC (*Sephadex LH-20*, $\text{CHCl}_3/\text{MeOH}$ 1 : 1) **1** (15 mg). Fr. **4** (21.6 g) was subjected to repeated CC ($\text{CHCl}_3/\text{MeOH}$ 50 : 1 \rightarrow 5 : 1), the obtained **2** was purified by CC (*Sephadex LH-20*, MeOH): **2** (25 mg).

2-(3,4-Dihydroxyphenyl)-3,5,10,11-tetrahydroxy-6-methyl-4H,8H-[2]benzopyrano[3,4-h]-1-benzopyran-4,8-dione (**1**): Yellow amorphous powder. UV (MeOH): 254 (3.52), 345 (3.10). IR (KBr): 3427, 1710, 1643, 1609, 1523, 1361, 1288, 1185, 1160, 794. ^1H - and ^{13}C -NMR: Table. ESI-MS: 449 ($[M - \text{H}]^-$). HR-ESI-MS: 449.0527 ($[M - \text{H}]^-$, $\text{C}_{23}\text{H}_{15}\text{O}_{10}$; calc. 449.0514).

2-(3,4-Dihydroxyphenyl)-3,5,11,12-tetrahydroxy-6-methyl-4H,8H-[2]benzopyrano[3,4-h]-1-benzopyran-4,8-dione (**2**): Yellow amorphous powder. UV (MeOH): 252 (3.50), 343 (3.11). IR (KBr): 3397, 1714, 1637, 1602, 1527, 1346, 1289, 1190, 1148, 795. ^1H - and ^{13}C -NMR: Table. ESI-MS: 449 ($[M - \text{H}]^-$). HR-ESI-MS: 449.0498 ($[M - \text{H}]^-$, $\text{C}_{23}\text{H}_{15}\text{O}_{10}$; calc. 449.0514).

REFERENCES

- [1] State Administration of Traditional Chinese Medicine, 'Zhong Hua Ben Cao', Shanghai Science and Technology Press, Shanghai, 1999, Vol. 5, p. 626.
- [2] I. Jantan, A. S. Ahmad, S. A. A. Bakar, A. R. Ahmad, M. Trockenbrodt, C. V. Chak, *Flavour Fragrance J.* **1998**, *13*, 245.
- [3] W.-Y. Tsui, G. D. Brown, *J. Nat. Prod.* **1996**, *59*, 1084.
- [4] Y. Fujimoto, S. Usui, M. Makino, M. Sumatra, *Phytochemistry* **1996**, *41*, 923.
- [5] W.-Y. Tsui, G. D. Brown, *Phytochemistry* **1996**, *43*, 871.
- [6] M. Makino, Y. Fujimoto, *Phytochemistry* **1999**, *50*, 273.
- [7] T. Satake, K. Kamiya, Y. Saiki, T. Hama, Y. Fujimoto, H. Endang, M. Umar, *Phytochemistry* **1999**, *50*, 303.
- [8] K. Kamiya, T. Satake, *Fitoterapia* **2010**, *81*, 185.
- [9] T. H. Quang, N. X. Cuong, C. V. Minh, P. V. Kiem, *Nat. Prod. Commun.* **2008**, *3*, 755.
- [10] E. Quezada, L. Santana, E. Uriarte, *Magn. Reson. Chem.* **2006**, *44*, 644.
- [11] L. Larsen, D. H. Yoon, R. T. Weavers, *Synth. Commun.* **2009**, *39*, 2935.
- [12] M. Yoshikawa, F. Xu, T. Morikawa, K. Ninomiya, H. Matsuda, *Bioorg. Med. Chem. Lett.* **2003**, *13*, 1045.
- [13] B. A. Bohm, 'Introduction to Flavonoids', Harwood Academic Publishers, Australia, 1998.
- [14] J.-T. Fan, Y.-S. Chen, W.-Y. Xu, L. Du, G.-Z. Zeng, Y.-M. Zhang, J. Su, Y. Li, N.-H. Tan, *Tetrahedron Lett.* **2010**, *51*, 6810.

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